

Tetrahedron Letters 43 (2002) 7721-7723

A convenient and highly stereoselective approach for α-galactosylation performed by galactopyranosyl dibenzyl phosphite with remote participating groups

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Received 9 April 2002; revised 22 August 2002; accepted 30 August 2002

Abstract—By using 4- and 6-acyl remote group participation, a galactosyl phosphite donor exhibits exceptionally high α -anomeric selectivity for primary and secondary hydroxyl acceptors. It was observed that temperature played an important factor in glycosylation. The higher α -selectivities were obtained at higher reaction temperatures. © 2002 Elsevier Science Ltd. All rights reserved.

Although numerous complex carbohydrates occur in nature,^{1,2} saccharide structures and functions have been minimally studied, mainly due to the difficulty of synthesizing oligosaccharides.^{3–7} The synthesis of 1,2-*cis* α -glycosides and β -mannopyranosides,⁸ for example, still represents a significant challenge. Since many carbohydrate antigens, such as α -galactosyl ceramide (Gal α 1-Ceramide, α -GalCer),⁹ α -Gal epitope (Gal α (1 \rightarrow 3)Gal), and P^k antigen (Gal α (1 \rightarrow 4)Gal), contain a 1,2-*cis*- α -galactosyl glycosidic bond, the development of new and more efficient methods to synthesize 1,2-*cis* glycosidic bonds is of current interest.

It has been shown^{6,10–12} that protecting groups on sugars affect the reaction rate and stereochemical outcome of glycosylation. Although highly α -selective galactosylation using a thiol galactosyl donor with remote electron-donating group participation at C-4 has been

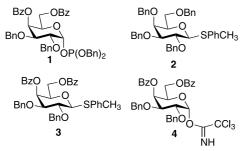


Figure 1. Galactosyl donors used in this study.

reported,^{13,14} a complicated solvent system was needed to ensure good selectivities.13 In addition, enhanced α -selectivity in galactosylation was achieved by employing the 6-O-acetyl participation strategy and phosphite leaving group.¹⁵ In certain cases, glycopyranosyl phosphites also showed better α -selectivity in glycosylation,16,17 but using commercially unavailable 2,6-di-tert-butylpyridinium iodide (DTBPI) as the promotor. As part of our continuing efforts in the synthesis of carbohydrate antigens as killer T-cell activators, we found that the 2,3-O-dibenzyl-4,6-O-dibenzoyl galactosyl phosphite donor 1 (Fig. 1) served as an excellent α -selective galactosyl donor under standard glycosylation reaction conditions. Compound 1 contains 4- and 6-acyl groups which may exhibit remote group participation, through the intermediates shown in Fig. 2, resulting in high α -anomeric selectivity.

Since aryl thiol glycosides are commonly used as α selective donors, we first examined glycosylation of acceptor **5** using **2** (Fig. 3). The reaction gave a good yield but low selectivity (Table 1, entry 2). In order to increase the α -selectivity, we introduced the remote group participation strategy and changed the protecting groups of the 4- and 6-hydroxyl groups of **2** to the benzoyl group (derivative **3**). Consequently, the replacement showed a significant improvement of α -selectivity from $\alpha:\beta=2:1$ to 9:1 (entry 3). To investigate further whether the leaving group at the anomeric center would influence the α -selectivity, the thiocresol group of **3** was replaced by trichloroimidate and dibenzyl phosphite to give **4** and **1**, respectively. Surprisingly, coupling of an

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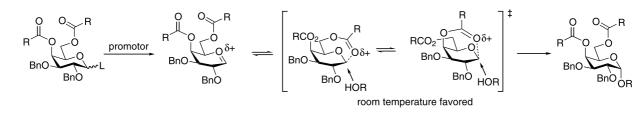
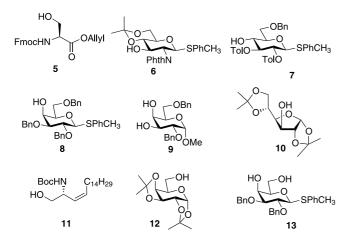


Figure 2. Proposed 4- and 6-acyl remote participation.

 α/β mixture ($\alpha:\beta=6:1$) of the phosphite donor 1 with 5 gave only the α -glycosylation product (entry 1), while using the imidate donor 4 (the α form) gave a mixture of products ($\alpha:\beta=1:10$, entry 4). The results of entries 1 and 4 showed that the glycosylation mechanism of phosphite donor 1 was $S_N 1$ like while that of imidate donor 4 was more $S_N 2$ like. These results encouraged us to investigate further the scope and limitation of phosphite donor 1 using a number of acceptors (Fig. 3).

As shown in Table 1, for secondary hydroxyl acceptors, phosphite donor 1 provided excellent α -selectivities (entries 5–9). It is worth pointing out that the 3-OH is more reactive than the 4-OH in the galactose case, thus we obtained disaccharide 18 in high yield and selectivity. However, when primary hydroxyl acceptors were used, the α -selectivity decreased, except with acceptor 5 (entries 1, and 10–12). The good α -selectivity of the primary acceptor 5 may be a result of an anomeric effect together with the π - π interaction between the 2-Bn of 1 and the Fmoc of 5 resulting in a nucleophilic approach from the α -face to give an α -glycosidic bond. This phenomenon has also been observed in the cases of fucosylation of tribenzyl fucosyl phosphite with Fmoc-Thr-OBzl.^{15,18} It should be noted that all the above mentioned glycosylations were performed at -15°C except those of entries 11 and 12. Interestingly, the selectivity of the galactosylation using donor 1 was temperature dependent. The higher α -selectivities were obtained at higher reaction temperatures. The temperature effect of entry 11 implied that the nucleophilic

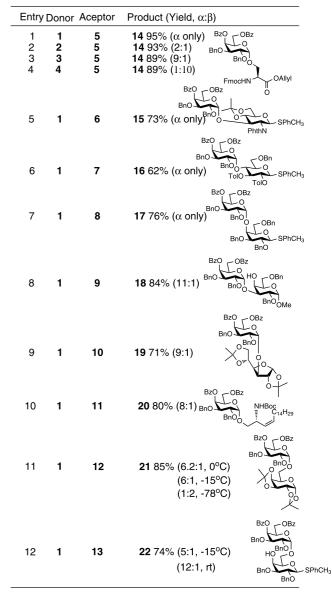


Tol=p-methylbenzoyl

Figure 3. Glycosylation acceptors used in this study.

addition to the phosphite donor becomes more like kinetic control at low reaction temperatures (-78° C), resulting in inversion of the reaction center for the major product. When the temperature is increased, the remote group participating effect is enhanced and thus gives better α -selectivity. The temperature effect was further demonstrated by running the glycosylation at

| Table | 1. ' | a |
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^a Tol=p-methylbenzoyl

room temperature as shown in entry 12. In comparison with the -15° C reaction, the α -selectivity at room temperature is significantly increased. Since the thiocresol leaving group is present on the disaccharides **15–17**¹⁹ and **21**, they can serve as disaccharide donors for the next glycosylation. Compound **17** was obtained in excellent α -selectivity and could be used as a synthon of P^k and Globo H antigens. Compounds **20** and **14**¹⁹ can be used as precursors for the synthesis of NK T cell activators, α -GalCer^{9,20,21} and its analogs.

In brief, we have demonstrated that galactosyl phosphite **1** is an excellent donor for α -galactosylation. Its superior selectivity may be due to 4- and 6-acyl group participation and π - π interaction between the acceptor and the donor. Glycosylation with this donor for the synthesis of NK T cell activators is underway.

General procedure for glycosylation

A mixed solution of 1 (1.33 g, 1.63 mmol) and 5 (0.55 g, 1.48 mmol) in CH₂Cl₂ (20 mL) in the presence of molecular sieves (2.4 g) was stirred at rt for 30 min, and then cooled to -15° C for another 30 min. TfOH (25.9 µL, 0.30 mmol) was added, and the mixture was stirred at -15° C under argon. After the acceptor disappeared on TLC, the mixture was filtered; the filtrate was concentrated and the residue was partitioned between ethyl acetate and NaHCO₃ solution. The aqueous layer was extracted with ethyl acetate. The combined organic layer was dried over MgSO₄, and concentrated. The resulting residue was checked by NMR to determine the ratio of α and β anomers, and purified on a silica gel column to give 95% yield of the product.

Acknowledgements

We thank Academia Sinica and the National Science Council (NSC 90-2323-B-001-005) for their generous support of this research, and Professor C.-H. Wong at The Scripps Research Institute for helpful discussions.

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- 19. Selective data: for compound 14: ¹H NMR (CDCl₃, 400 MHz) δ 3.94 (dd, J=3.6, 10.0 Hz, 1H), 3.98 (dd, J=3.2, 10.0 Hz, 1H), 4.08 (dd, J=3.2, 10.0 Hz, 1H), 4.10-4.20 (m, 2H), 4.26–4.39 (m, 4H), 4.50 (dd, J=7.2, 11.2 Hz, 1H), 4.54–4.62 (m, 3H), 4.62 (d, J=12.0 Hz, 2H), 4.76 (d, J=12.0 Hz, 2H), 4.85 (d, J=11.2 Hz, 1H), 4.92 (d, J = 306 Hz, 1H), 5.17 (dd, J = 1.2, 10.4 Hz, 1H), 5.27 (dd, J = 1.2, 17.2 Hz, 1H), 5.79–5.90 (m, 1H), 5.89 (d, J = 3.2Hz, 1H), 6.09 (d, J=8.4 Hz, 1H), 7.20-7.62 (m, 22H), 7.50 (d, J=8.0 Hz, 2H), 7.97-8.07 (m, 4H); compound 16: ¹H NMR (CDCl₃, 400 MHz) δ 2.27 (s, 3H), 2.32 (s, 3H), 2.35 (s, 3H), 3.62 (dd, J=3.2, 10.4 Hz, 1H), 3.63-3.75 (m, 2H), 3.93 (dd, J=3.2, 10.4 Hz, 1H), 3.96 (dd, J = 3.6, 11.2 Hz, 1H'), 3.97 (d, J = 12.4 Hz, 1H), 4.17 (dd, J=5.2, 11.2 Hz, 1H), 4.22 (t, J=9.6 Hz, 1H), 4.23 (d, J=12.4 Hz, 1H), 4.32 (d, J=10.8 Hz, 1H), 4.34 (dd, J=6.4, 11.2 Hz, 1H), 4.40 (t, J=6.4 Hz, 1H), 4.51 (s, 2H), 4.66 (d, J = 10.8 Hz, 1H'), 4.82 (d, J = 10.0 Hz, 1H'), 5.02 (d, J = 3.2 Hz, 1H), 5.37 (t, J = 9.6 Hz, 1H'), 5.72 (m, 1H), 5.74 (t, J=9.6 Hz, 1H'), 7.00–7.60 (m, 29H), 7.82 (dd, J = 6.8, 8.0 Hz, 4H), 7.87 - 8.03 (m, 4H).
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